Commonly used *Streptomyces* media

Medium	**Ingredients and notes**
Solid Minimal Medium (Kieser *et al.*, 2000))	0.5 g or 1 g L-asparagine or $(NH_4)_2SO_4$ 0.5 g K_2HPO_4 0.2 g $MgSO_4*7H_2O$ 0.01 g $FeSO_4$ *7 H_2O 10 g Agar Add 1000 ml dH_2O, pH to 7.0 with 5M NaOH, Make up 1% carbon (see below) and add after autoclaving.
Solid Minimal Medium (Karandikar *et al.*, 1997)	1.7 g $NaNO_3$ 1 g K_2HPO_4 0.5 g KCl 0.5 g $MgSO_4*7\ H_2O$ 0.01 g $FeSO_4*7\ H_2O$ 2.5 mL/L Trace element solution * 15 g Agar Add 800 ml dH2O, pH to 7.0 with 5M NaOH, add 200 ml of 50 g/l glucose after autoclaving.
Hobbs Minimal Medium (Hobbs *et al.*, 1989)	5 g NaCl 5 g Na_2SO_4 4.5 g $NaNO_3$ 1.2 g TRIS 2 g $MgSO_4*\ 7\ H_2O$ 0.01 g $ZnSO_4*\ 7\ H_2O$ Add 1000 ml dH_2O, pH was adjusted to 7.2 added after autoclaving: 1ml K_2HPO4 (20 g/L stock) 1 mL/L trace element solution * either glucose or Tween40 were used as carbon source (0.33 mol carbon) 20 µL of Sigma Antifoam A

Mannitol Soya Flour Medium (Hobbs *et al.*, 1989)	16 g Mannitol 16 g Soya Flour 16 g Agar Add 1000 ml tap water Make in 250 ml batches and autoclave in 500 ml Duran bottles as the medium bubbles out
R2 (Solid) (Kieser *et al.*, 2000)	103 g Sucrose 0.25 g K_2SO_4 5 g $MgCl_2$ *6 H_2O 10 g glucose 0.1 g casein hydrolysate 2 mL/L trace element solution ** 5.73 g TES buffer 22 g agar Add 1000 ml dH_2O (heat to dissolve the components and aliquot in to 200 ml batches for autoclaving) additional components added after autoclaving (per 100 mL) 1 mL 0.5% KH_2PO_4 0.4 mL 3.68% $CaCl_2$*2 H_2O 1.5 mL 20 % L-Proline 0.7 mL 1 M NaOH

R5 (Solid) (Kieser *et al.*, 2000)	103g sucrose 0.25 g K_2SO_4 10.12 g $MgCl_2$*6 H_2O 10 g glucose 0.1 g casein hydrolysate 2 mL/L trace element solution** 5 g yeast extract 5.73 g TES buffer 27.5 g agar Add 1000 ml dH_2O (heat to dissolve the components and aliquot in to 320 ml batches for autoclaving). Additional components added after autoclaving (per 80 mL) 1 mL 0.5% KH_2PO_4 0.4 mL 5 M $CaCl_2$*2 H_2O 1.5 mL 20% L-Proline 0.7 mL of 1 M NaOH
2xYT (Liquid) (Kieser *et al.*, 2000)	16 g tryptone 10 g yeast extract 5 g NaCl Add 1000 ml dH_2O
YEME (Liquid and solid) (Hoskisson *et al.*, 2000; Kieser *et al.*, 2000)	5 g peptone 3 g yeast extract 3 g malt extract 10 g glucose **OPTIONAL:** 340 g sucrose (Can be added for more dispersed growth). Add 1000 ml dH_2O – Adjust pH to 7.2 with 5 M NaOH. Make in 250 ml batches and autoclave in 500 ml Duran bottles as the medium can bubble out. A solid version of this can be made by omitting the sucrose and adding 20 g of agar per litre. Again, make in 250 ml batches and autoclave in 500 ml Duran bottles as the medium bubbles out.

* Trace element solution of Hobbs et al., (1989) consists of (g/100 mL; Filter sterilised): 0.1 $ZnSO_4$*7 H_2O, 0.1 $FeSo_4$ *7 H2O, 0.1 $MnCl_2$ * 4 H_2O, 0.1 $CaCl_2$ * 6 H_2O and 0.1 NaCl, can be stored at 4◦ C for 2−4 weeks.It is diluted 1:10 before added to the medium

** Trace element solution was composed of: 8.78 g/L $FeCl_3$, 2.04 g/L $ZnSO_4$, 1.02 g/L $MnCl_2$*4 H_2O, 0.43 g/L $CuSO_4$*2 H_2O, 0.42 g/L NaI, 0.31 g/L H_3BO_3, 0.24 g/L $CaCl_2$*6 H_2O and 0.24 g/L Na_2MoO_4*2 H_2O

Carbon sources added to minimal medium for screening

Carbon source	Stock/pH	mL to 1L medium (1 %)
Glucose	50	20
Mannitol	25	40
Tween80	20	50
Pyruvate	20/ pH 7.2	50
Na-Acetate	20/ pH 7.2	50
Na-Citrate	20/ pH 7.2	50
Malic acid	20/ pH 7.2	50
PEP	20	50
α-KG	20	50
GlcNac	20	50

References

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